

# FACTSHEETS OF PESTS OF PHYTOSANITARY SIGNIFICANCE TO KENYA

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# Helicoverpa armigera

# Identity

Preferred Scientific Name: Helicoverpa armigera (Hübner, 1809)

Other Scientific Names: Bombyx obsolete Fabricius, Chloridea armigera Hübner, Chloridea obsoleta Fabricius, Helicoverpa commoni Hardwick, Helicoverpa obsolete Auctorum, Heliothis armigera (Hübner), Heliothis conferta Walker, Heliothis fusca Cockerell, Heliothis obsoleta Auctorum, Heliothis pulverosa Walker, Heliothis rama Bhattacherjee Gupta, Heliothis uniformis Wallengren, Noctua armigera Hübner and Noctua barbara Fabricius.



Helicoverpa armigera

**Preferred Common Name:** Cotton bollworm, Corn earworm, African bollworm, Gram pod borer, old world bollworm or tobacco budworm.

Taxonomic Position: Class: Insecta, Order: Lepidoptera, Family: Noctuidae

# Hosts/species affected

It is a primarily pest of ornamental plants e.g roses, flowers, vegetable crops, cotton, tomatoes, Potatoes, Maize, Soya beans, lucerne, chickpeas, sorghum, *Phaseolus*, fruits (*Prunus*, *Citrus*) and forest trees.

# Growth stages of host plant affected

- Vegetative growing stage
- Flowering stage
- Fruiting stage.



# Biology and Ecology

Eggs: Eggs are 0.5 mm in diameter, round and yellowish-white in colour. The eggs darken before hatching and are deposited singly on tender parts of the plant. Egg-laying cycle generally coincides with early flowering of host crops.

Larvae: Larvae are yellowish-white to reddish brown in colour. Dark brown to black head, rows of black bumps with short hairs along the backs giving spotted appearance. 35 - 40 mm long when fully grown and normally drop from the plant and burrow into the soil to pupate.



Figure 1. Yellowish-white eggs before hatching. Photo: A.M. Varela, IC IPE.

Pupa: Pupa is shiny brown, about 16 mm long, smooth surface with two short parallel spines at the posterior tip of the body.

Adult: Adults are yellowish-brown with a dark speck, greyish irregular lines and a black kidney-shaped mark on the forewings. Hindwings are whitish with a black patch along the outer margin. The adults are 14 to 18 mm long with a wingspan Figure 2. Fully grown caterpillars of 35 to 40 mm.



3-4 cm long. Photo: A.M. Varela, ICIPE.

Figure 3. Shiny brown, about 16 mm long pupa. Photo: A.M. Varela, ICIPE.



Figure 4. Dark speck on the forewings of adult month. Photo: A.M. Varela, ICIPE



# Life cycle:

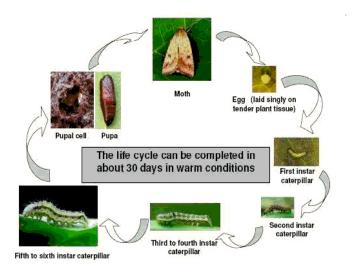


Figure 5. Life cycle is completed in about 25 to 60 days depending on temperature. Courtesy of A.M. Varela, ICIPE.

# Symptoms

Larvae feed on leaves, flower buds, flowers, developing grains and bore into pods and fruits. The larvae also deposits excrements on damaged plant parts.

# Means of movement and dispersal

Adults can migrate over long distances. Movement in international trade is mainly on ornamental plants and on cut flowers, in cotton bolls and tomato fruits.

## **Impact**

H. armigera cause serious losses in particular to cotton, tomatoes and maize. On cotton, two to three larvae on a plant can destroy all the bolls within 15 days. On maize and baby corn, they consume developing grains. On tomatoes, they invade fruits, preventing development and causing falling. It destroys the quality of ornamental plants by feeding on leaves and inflorescence.

#### Movement in trade

The plant parts liable to carry the pest in trade include: Fruits, leaves and inflorescence.



## **Detection and inspection**

The feeding larvae can be seen on the surface of plants but they are often hidden within plant organs (flowers, fruits etc.). Feeding holes and heaps of frass (excreta) are visible, but it is necessary to cut open the plant organs to detect the pest.

# Phytosanitary significance

In the EU directive 2000/29/EC, Annex 1 part A, the pest is listed in section II as a harmful organism known to occur in the community and relevant for the entire community (EPPO, 2016).

#### Management

# Cultural practices

## • Monitoring and Decision Making:

Early detection is achieved by regular scouting of the crop. Monitoring moth population can also be done by use of pheromone traps.

#### • Sanitation

- > Remove and destroy plant residues immediately after harvesting.
- ➤ Plough the soil after harvesting. This exposes pupae, which may then be killed by natural enemies or through desiccation by the sun.

# • Mechanical control

- ➤ Hand picking and destroy eggs and small caterpillars. This is feasible in small plots or green houses when infestation is low.
- > Sort out the harvested crop very thoroughly during grading to remove the caterpillars manually.

# Habitat Management

## • Intercropping and trap crops

Adult Moths prefer laying eggs on certain crops (e.g. pigeon pea, chick pea, crotalaria, maize, tobacco, African marigold, sorghum and sunflower), especially during the flowering period. Those crops may be utilized to distract moths from crops that are more vulnerable to African bollworm damage. If trap crops are planted in strips or around the field, the moths will lay eggs on them instead of the main crop. African marigold has been used as trap crop in tomato. Marigold planted after every eight rows of tomato has been observed to attract most of the African bollworms moths to marigold (Negash and Abate, 2002).



Intercropping of cotton with chickpea, cowpea, onion, pearl millet, crotalaria, pigeon pea and marigold in strips is reported to divert the populations of sucking pests and the African bollworm from cotton (Dejen and Tesfaye, 2002).

# • Crop rotation

Avoid planting crops after each other that are susceptible to bollworm like cotton, maize, sorghum, tobacco, soybean and tomato. This helps in preventing build-up of bollworm populations. To be effective crop rotation should be done over large areas since moths can fly long distances (CABI, 2002).

Some crops not susceptible to bollworm that could be planted in rotation with susceptible crops are small grains like rice and plants of the onion family.

## **Biological Control**

• Natural enemies: Trichogramma spp has been tried successfully as an egg parasitoid (Cherry et al, 2003).

#### Chemical Control

Various chemical and Bio-pesticide products are registered in Kenya against *H. armigera* available in various brand names.

Name of product (Active Ingredient)	Purpose of registration
Dimethoate 400 g/L, Imidacloprid 100 g/L +	Control of H. armigera in Cotton
Betacyfluthrin 45g/L, Cyperm ethrin 10% w/v +	
Chlorpyrifos 35% w/v and Alpha-cypermethrin 10g/L	
Indoxacarb 150g/L	Control of H. armigera in
	Tomatoes
Bacillus Thuringiensis var. Kurstaki (Btk) 9 x 107	Control of H. armigera in French
spores/m g	beans
Helicoverpa armigera SNPV 8% w/w 2x99 polyhedra per	Control of African bollworm in
ml, Etofenprox 30% and Tebufenoxide 20%	Tomatoes
Methomyl 90% w/w	Control of H. armigera in Flowers
	and horticultural crops.

For more information refer to the list of registered pest control products provided by the Pest Control Products Board (PCPB) Website: www.pcpb.or.ke

Note: Repeated use of a given chemical creates resistance.



#### References

- CABI. (2016). Crop Protection Compendium, 2016 Edition. © CAB International Publishing. Wallingford, UK. www.cabi.org. Retrieved on 23<sup>th</sup> April 2016
- Cherry, A., Cock, M, van den Berg, and Kfir, R (2003). Biological control of *Helicoverpa* armiguera in Africa. In Biological Control in IPM Systems in Africa. Neuenschwander, P., Borgemeister, C and Langewald. J. (Editors). CABI Publishing in association with the ACP-EU Technical Centre for Agricultural and Rural Cooperation (CTA) and the Swiss Agency for Development and Cooperation (SDC). pp. 329-346. ISBN: 0-85199-639-6.
- Dejen, A. and Tesfaye (2002). Cultural control of the African bollworm, *Heliothis armiguera* (Hb.). In African Bollworm Management in Ethiopia. Status and needs. Proceedings of the National Workshop held at the Plant Protection Research Centre Ambo, Ethiopia. April 2002. pp 99-105.
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- Negash, T. and Abate, S. (2002). African bollworm management in the southern Region of Ethiopia. In African Bollworm Management in Ethiopia. Status and Needs. Proceedings of the National Workshop held at the Plant Protection Research Centre Ambo, Ethiopia. April 2002. pp 59-62.

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